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Taxonomic re-evaluation of the Taiwanese montane earthworm *Amynthas wulinensis* Tsai, Shen & Tsai, 2001 (Oligochaeta: Megascolecidae): Polytypic species or species complex?

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Abstract

Body size and colouration are two characters commonly used in the taxonomy of many animal taxa. However, they are seldom used by earthworm taxonomists because they are subject to environmental influences and tend to vary intraspecifically. In the present study, DNA sequences of the mitochondrial COI gene are used to evaluate whether specimens of the megascolecid earthworm *Amynthas wulinensis* Tsai, Shen & Tsai, 2001 that differ in body size and/or colouration belong to different genetic lineages. Phylogenetic analyses and morphological comparisons indicate that *A. wulinensis* in the previous broad sense is a species complex composed of three species differing in body size, colouration, and genital markings. Consequently, two new species, *Amynthas lini* and *Amynthas meishanensis*, are described. Taxonomic affinities of the *A. wulinensis* species complex are discussed, as is the feasibility of using body size and colouration in earthworm taxonomy.

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Introduction

Morphological characters, both quantitative and qualitative, are used by morphological taxonomists for species identification. In earthworm systematics, reproductive organs, including the clitella, male pores, female pores, testes, ovaries, spermathecae, and prostate glands, are given the greatest importance, as these characters are believed to be less affected by environ-

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mental fluctuations through time, and are evolutionarily more conservative. Other characters used by earthworm taxonomists include some anatomical features, e.g., the alimentary canal, nephridia, nervous system, and vascular system. Moreover, some external characters, such as the prostomia, dorsal pores, setae, and genital markings, are also valuable in species identification (Gates 1972; Blakemore 2002). Besides these characters, body size and colouration are two external characters commonly and extensively used in the taxonomy of many animal taxa. However, in earthworms these two characters are evolutionarily less conservative, subject to environmental influence, and intraspecifically variable. In addition, they are easily affected by the methods of

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sample processing and preservation. That is why the use of body size and colouration has been limited in earthworm species identification (Gates 1972; Blakemore 2002).

Amynthas wulinensis Tsai, Shen & Tsai, 2001, a member of the pheretimoid earthworms within the Megascolecidae, is an octothecate montane earthworm reported from central Taiwan. Its body size, colouration, and genital markings vary among different collection sites and elevations. The 63 specimens reported from central Taiwan have variable postclitellar genital papillae but no preclitellar ones (Tsai et al. 2001). Their body length varies from 6.3 cm in some specimens collected at lower elevations to 17.4 cm in some collected at higher elevations (Tsai et al. 2001). The species may be more variable still: some newly collected specimens from northern and central Taiwan have lengths exceeding 20 cm, greater than those reported in the original description, and most of them, though not all, have preclitellar genital papillae (H.-P. Shen personal communication; authors' personal observation). Additional newly collected specimens from south-central Taiwan are similar to those reported from Nanshan Creek, Mt. Beidongyan, and Meifeng (Tsai et al. 2001), but have preclitellar genital papillae (authors' personal observation). Moreover, most, if not all, of the large live specimens, i.e., specimens with body lengths exceeding 20 cm, show quite dark pigmentation on the setal annuli, resulting in a striped appearance. This unique body colouration has never been observed in smaller live specimens. Collectively, these differences in body size, colouration, and genital markings may be the result of environmental variation in different habitats. Alternatively, specimens with different morphological character states may belong to different genetic lineages or even different species. Further evaluation of this problem requires information from studies based on molecular approaches, such as DNA sequence analyses.

Among the published papers, only four earthworm studies focused on genetic differentiation between congeneric species or between different populations of a single species using DNA sequence analyses. Jamieson et al. (2002) used rDNA data to separate some congeneric species of Megascolecidae. Heethoff et al. (2004) used cytochrome c oxidase subunit II (COII) gene sequences to analyse the genetic differentiation between two parthenogenetic populations of the lumbricid earthworm Octolasion tyrtaeum (Savigny, 1826) differing in body size. Pérez-Losada et al. (2005) used 28S rRNA and COI gene sequences to verify that two earthworms, Eisenia fetida (Savigny, 1826) and E. andrei (Bouché, 1972), are both valid phylogenetic species. Chang and Chen (2005) used cytochrome c oxidase subunit I (COI) gene sequences to re-evaluate the taxonomic status of two sibling pheretimoid earthworms, Metaphire formosae (Michaelsen, 1922) and

M. yuhsii (Tsai, 1964). They also analysed the genetic differentiation between two different populations of M. yuhsii. These four studies show that molecular approaches are applicable and very useful to research on the taxonomy and intraspecific variation of earthworms. In the present study, we attempt to re-evaluate the taxonomic status of A. wulinensis with the aid of DNA sequence analysis. We also discuss the feasibility of applying body size and colouration in species identification.

Materials and methods

Sample collection and preservation

Samples of *A. wulinensis* were collected from various localities in Taiwan. The earthworms were anesthetised in a 10% ethanol solution. Some muscle tissues were isolated and preserved in a 70% or 95% ethanol solution for DNA extraction. The residual earthworm samples were fixed in 10% formalin and preserved in a 70% ethanol solution. Some other *Amynthas* species collected in Taiwan were treated using the same procedure for the phylogenetic analysis. The earthworm specimens are deposited at the Institute of Zoology, National Taiwan University, Taipei.

DNA extraction, PCR, and DNA sequencing

Muscle tissues were washed with distilled water, homogenised in liquid nitrogen, and digested in digestion buffer ($10\,\text{mM}$ Tris–HCl, $2\,\text{mM}$ dihydrate EDTA, $10\,\text{mM}$ NaCl, $10\,\text{mg/ml}$ DTT, 1% SDS, and $0.4\,\text{mg/ml}$ proteinase K) at $50\,^{\circ}\text{C}$ for $15-30\,\text{min}$. Total DNA was extracted from the digested tissue-buffer solution with a standard phenol/chloroform extraction method (Palumbi et al., 1991). The ethanol-precipitated DNA was dissolved in distilled water, checked with 1.0% agarose gel electrophoresis, and stored at $-20\,^{\circ}\text{C}$.

A 626-bp mitochondrial cytochrome *c* oxidase subunit I (COI) DNA fragment was amplified using the universal primers LCO1490 (5'-GGT CAA CAA ATC ATA AAG ATA TTG G-3') and HCO2198 (5'-TAA ACT TCA GGG TGA CCA AAA AAT CA-3') (Folmer et al. 1994). PCR amplifications were carried out in a 50 μl total volume, using 1 cycle at 94 °C for 1 min, followed by 35 cycles of denaturation for 30 s at 94 °C, annealing for 30 s at 54 °C, and extension for 50 s at 72 °C, with a final extension at 72 °C for 10 min.

The PCR products were checked using 1.0% agarose gel electrophoresis and sequenced in both directions using the same primers as in the PCR. Sequencing was performed with the ABI PRISM BigDye Terminator Cycle Sequencing Ready Reaction Kit, V3.1 (Applied Biosystems, CA, USA). Products were analysed with a

ABI 3730 XL DNA analyzer (Applied Biosystems). The computer base calling was double-checked by eye.

Sequence alignment and phylogenetic analyses

The sequence was checked by aligning the sequencing results with the COI full sequence of *Lumbricus terrestris* (GenBank accession no. U24570) using the default settings of Clustal X 1.81 (Thompson et al. 1997).

The sequence of *Pontodrilus litoralis* was retrieved from GenBank (Table 1) and used as an outgroup in the phylogenetic analyses. All sequences analysed were aligned using the default settings of Clustal X 1.81. A homologous fragment of 626 bp of the COI sequence was used in this study. The sequences obtained were submitted to GenBank (see Table 1). Neighbour-joining (NJ) analyses were performed using MEGA 2.1 (Kumar et al. 2001) with Kimura's (1980) two-parameter model. Maximum-parsimony (MP) analyses were performed using PAUP 4.0b10 (Swofford 2000) with heuristic searches, random starting trees, 100 random additions of sequences, and TBR branch swapping. Bootstrapping

with 1000 pseudo-replicates for the NJ and MP analyses was used to examine the robustness of the clades and their phylogenetic relationships.

Morphological comparisons

The characters described in the original description of *A. wulinensis* (Tsai et al. 2001), such as body length, clitellum width, spermathecae, male pores, testes, setae, and genital markings, were used to compare the intraspecific clades revealed in the phylogenetic analyses.

Results

Phylogenetic analyses

The samples used in the phylogenetic analyses and the corresponding GenBank accession numbers of the COI sequences are listed in Table 1. In the 626-bp COI

Table 1. Samples used in the phylogenetic analyses, and corresponding GenBank accession numbers; asterisks indicate sequences retrieved from GenBank

OTUs	Locality	Sample no.	Accession no.
Amynthas wulinensis s.l. (clade A)	Wulai, Taipei County	B0415	DQ224166
	Wulai, Taipei County	B0416	DQ224167
	Wulai, Taipei County	B0417	DQ224168
	Wulai, Taipei County	B0418	DQ224169
	Meishan, Chiayi County	B2802	DQ224170
	Meishan, Chiayi County	B2803	DQ224171
	Meishan, Chiayi County	B2804	DQ224172
	Meishan, Chiayi County	B2805	DQ224173
	Meishan, Chiayi County	B2806	DQ224174
A. wulinensis s.l. (clade B)	Kunyang, Nantou County	B2703	DQ224175
	Kunyang, Nantou County	B2704	DQ224176
	Kunyang, Nantou County	B2705	DQ224177
	Kunyang, Nantou County	B2706	DQ224178
	Kunyang, Nantou County	B2707	DQ224179
	Kunyang, Nantou County	B2708	DQ224180
	Meifeng, Nantou County	B2901	DQ224181
	Meifeng, Nantou County	B2902	DQ224182
A. wulinensis s.l. (clade C)	Meishan, Chiayi County	B2903	DQ224183
	Meishan, Chiayi County	B2904	DQ224184
	Meishan, Chiayi County	B2905	DQ224185
	Meishan, Chiayi County	B2906	DQ224186
	Meishan, Chiayi County	B2907	DQ224187
A. tayalis	Wulai, Taipei County		AY962185*
A. binoculatus	Baoshan, Hsinchu County		AY962184*
A. aspergillus	Lienhuachih, Nantou County		DQ224188
A. polyglandularis	Wulai, Taipei County		DQ224189
A. corticis	Taipei City		DQ224190
A. robustus	Taipei City		DQ224191
Pontodrilus litoralis			AF003256*

sequence, no insertions or deletions were found. From 22 individuals of *A. wulinensis*, 16 haplotypes of the COI gene were observed: seven, six, and three haplotypes belonged to clades A, B, and C in the phylogenetic analyses, respectively (see below). The genetic distances among different clades and other *Amynthas* species are listed in Table 2. The average genetic distances between each pair of clades A, B, and C of *A. wulinensis* were equivalent to the interspecific genetic distances between other *Amynthas* species. Genetic distances between any pair of individuals within each of clades A, B, and C were 0–14.6%, 0–9.9%, and 0–0.8%, respectively. All of these values were smaller than the genetic distances between each pair of the three clades.

For the 626-bp sequence, 244 variable sites were observed, among which 216 were parsimony informative. The phylogenetic analyses resulted in a strictconsensus tree from the three MP trees equalling 799 steps (Fig. 1), with a consistency index (CI) of 0.4731, a homoplasy index (HI) of 0.5269, a retention index (RI) of 0.7455, and a rescaled consistency index (RC) of 0.3527. The NJ tree had a topology and bootstrap values similar to the strict-consensus MP tree (Fig. 1). The two trees strongly indicate A. wulinensis sensu lato as being non-monophyletic, composed of three separate monophyletic clades A, B, and C (respective bootstrap values in the NJ and MP trees: 99 and 91 for clade A, 100 and 100 for clade B, 100 and 100 for clade C). The presence of three clades and the amounts of sequence divergence between them are clear evidence that specimens formerly identified as A. wulinensis belong to a species complex composed of three different species corresponding to clades A, B, and C.

Taxonomic section

According to the morphology and collection sites, clade B in the phylogenetic analyses is A. wulinensis Tsai

Table 2. Pairwise comparisons of *Amynthas wulinensis* and other *Amynthas* species in Taiwan by Kimura's two-parameter distance

OTUs	AwuA	AwuB	AwuC	Apo	Ata	Abi	Aas	Aro
AwuB	0.193							
AwuC	0.223	0.208						
Apo	0.209	0.208	0.197					
Ata	0.190	0.188	0.152	0.198				
Abi	0.204	0.204	0.198	0.208	0.198			
Aas	0.225	0.211	0.244	0.202	0.214	0.225		
Aro	0.186	0.179	0.214	0.220	0.205	0.201	0.169	
Aco	0.179	0.188	0.203	0.198	0.168	0.207	0.191	0.168

AwuA = clade A of A. wulinensis; AwuB = clade B of A. wulinensis; AwuC = clade C of A. wulinensis; Apo = A. polyglandularis; Ata = A. tayalis; Abi = A. binoculatus; Aas = A. aspergillus; Aro = A. robustus; Aco = A. corticis.

et al., 2001, whereas clades A and C are new species. The two new species are described below.

Amynthas lini sp. n. (Fig. 2)

Etymology

The species epithet is given in honour of the Taiwanese zoologist Dr. Yao-Sung Lin, who is promoting earthworm studies in Taiwan.

Material

Type material

Holotype: Mature (clitellate) specimen, dissected; Taiwan, Taipei Co., Wulia, 24 November 2001, C.-H. Chang (cat. no. 14-02815; Institute of Zoology, National Taiwan University, Taipei). Paratypes (all mature (clitellate) specimens, dissected): Two specimens; data as for holotype (14-02801 and 14-02814). One specimen; as holotype, except 1 August 2001 (14-02802). One specimen; as holotype, except 9 June 2000, C.-E. Lee (14-7292). One specimen; Taipei Co., Hsintien, 26 October 2000, S.-P. Wu (14-7291). One specimen; northern Taiwan, Ilan Co., Jiaushi, 18 May 2002, I.-H. Chen (14-03864). One specimen; northwestern Taiwan, Taoyuan Co., region surrounding Shihmen Reservoir, 14 December 2002, I.-H. Chen (14-05451).

Other material examined (all mature (clitellate) specimens)

One specimen (dissected); Taipei City, 9 August 2000, C.-H. Chang (14-00867). One specimen (dissected); Wulai, 1 August 1999, S.-P. Wu (14-00870). One specimen (dissected); central Taiwan, Nantou Co., Meifeng, 16 September 1999, I.-H. Chen (14-02463). Two specimens; Wulai, 1 August 2001, C.-H. Chang (14-02803 and 14-02805). One specimen; Jiaushi, 18 May 2002, I.-H. Chen (14-03885). One specimen; northcentral Taiwan, Hsinchu Co., Taoshan, 4 March 2003, I.-H. Chen (14-05889). One specimen; Hsinchu Co., Jiuchongping, 24 July 2003. I.-H. Chen (14-06552). One specimen; central Taiwan, Nantou Co., Kunyang, 22 October 2004, C.-H. Chang (14-07249). Two specimens; south-central Taiwan, Chiayi Co., Meishan, 6 December 2003, C.-H. Chang (14-07293 and 14-07294). One specimen (dissected); Meifeng, 5 October 2002, I.-H. Chen (14-07295). Two specimens; Meishan, 24 October 2004, C.-H. Chang (14-07254 and 14-07255).

External characters

Length 212–254 mm; clitellum width 7.0–9.0 mm; segment number 117–129; number of annuli per segment 1 in I–V, 3 in VI and beyond; prostomium epilobous; setae 20–26 in V, 33–45 in VII, 49–57 in X,

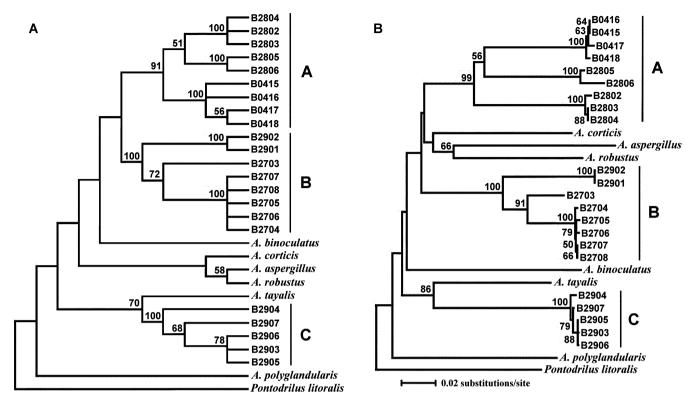


Fig. 1. Phylogenetic trees of *Amynthas wulinensis* and some congeneric species, based on DNA sequences of the mitochondrial COI gene. (A) Strict consensus of three maximum-parsimony trees reconstructed using heuristic searches, 100 random additions of sequences, and TBR branch swapping. (B) Neighbour-joining tree reconstructed using Kimura's two-parameter model. Bootstrap support values > 50 indicated at nodes. See text on clades (A), (B) and (C).

8–17 between male pores; first dorsal pore in 12/13; clitellum XIV–XVI, smooth; setae and dorsal pores absent.

Spermathecal pores four pairs in 5/6-8/9, ventrolaterally. Distance between paired spermathecal pores about 0.40 body circumference ventrally. Genital papillae present or absent. When present, papillae presetal, postsetal, or both. Presetal papillae 1–4 pairs in VII-X. Each papilla large, round, disc-like; distance between paired genital papillae about 0.10-0.30 body circumference ventrally. Sometimes only 1 papilla present on some segments. Postsetal papillae two pairs in VII and VIII, similar to presetal ones but smaller, distance between paired genital papillae about 0.40 body circumference ventrally. Sometimes only 1 papilla present on some segments. Female pore single, medioventrally in XIV. Male pores paired in XVIII, lateroventrally, distance between pores about 0.33 body circumference ventrally. Porophores round or oval on setal line, with a concave centre surrounded by 2 or 3 circular folds. Fold lines on some specimens parallel with setal line. Genital papillae postsetal, paired in XIX. Two specimens with an additional pair in XVII. One specimen with an additional pair in XX. Two specimens with 2 additional pairs in XVII and XX. Each papilla oval, with a concave centre surrounded by a few circular folds.

Preserved specimens dark brown on dorsum and around clitellum, light yellow on venter and setal lines, forming a striped or banded appearance, a dark brown circular band, and a light yellow one in sequence.

Internal characters

Septa 5/6–7/8 thickened, 8/9 and 9/10 absent, 10/11–13/14 greatly thickened. Gizzard round in X, almond-coloured. Intestine enlarged from XV. Intestinal caeca paired in XXVII, simple, surface slightly folded with the septa, grey-coloured, extending to XXIII or XXII. Lateral hearts enlarged in XI–XIV.

Spermathecae four pairs in VI–IX, with a short thick stalk about 0.45 mm long. Ampulla round, light yellow, about 3.00 mm in diameter. Spermathecal diverticula present, with a small oval seminal chamber of 2.07 mm and a slender, straight stalk of 1.69 mm. Nephridia tufted, attached to post-segmental septa, surrounding segmental chambers anterior to septum 6/7. No nephridia on spermathecal ducts. Ovaries paired in XIII, medioventral, close to septum 12/13.

Testes paired in X and XI, small, irregular, light yellow. Seminal vesicles paired in XI and XII, large, white or light brown. Prostate glands paired in XVIII, large, almond-coloured, extending anteriorly to XVI, with a

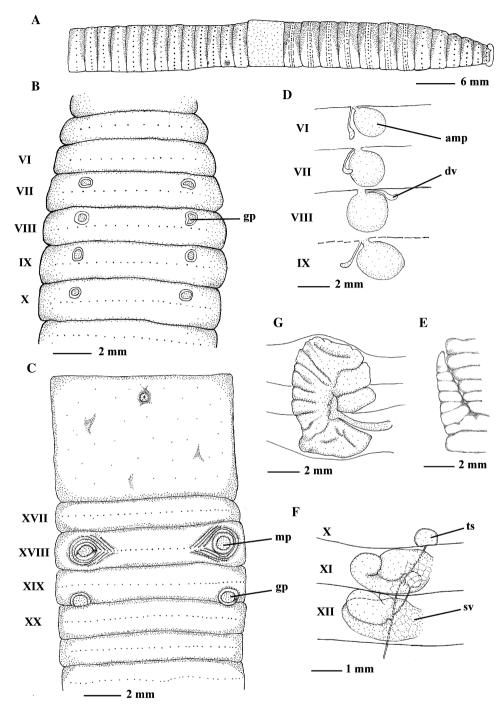


Fig. 2. Amynthas lini, morphology. (A) Lateral view of anterior body. (B) Ventral view of spermathecal pore region; gp = genital pad. (C) Male pore area; gp = genital pad, mp = male pore. (D) Left spermathecae; amp = ampulla, dv = diverticulum. (E) Left caecum. (F) Left testis sacs and seminal vesicles; ts = testis sac, sv = seminal vesicle. (G) Left prostate gland.

thick straight duct (Fig. 2). Accessory glands paired in XIX, with positions corresponding to external papillae.

the ground on rainy days and can often be found in drainage ditches along roads.

Localities and habitats

Amynthas lini sp. n. is distributed in mountainous areas of northern and central Taiwan, at elevations from 400 to 3000 m (Fig. 4). Animals usually crawl out onto

Amynthas meishanensis sp. n. (Fig. 3)

Etymology

The species epithet, to be treated as an adjective, refers to the type locality.

Material

Type material

Holotype: Mature (clitellate) specimen, dissected; south-central Taiwan, Chiayi Co., Meishan, 24 October 2004, C.-H. Chang and Y.-H. Lin (cat. no. 14-07290; Institute of Zoology, National Taiwan University, Taipei). Paratypes: Three mature (clitellate) specimens (one dissected); same data as holotype (14-07285, 14-07288, and 14-07289).

Other material examined

Ten mature (clitellate) specimens (7 dissected); same data as holotype (14-07258–14-07264, 14-07284, 14-07286, and 14-07287).

External characters

Length 38–65 mm; clitellum width 2.7–3.5 mm; segment number 51–113; one annulus per segment on all segments; prostomium epilobous; setae 27–31 in V, 35–42 in VII, 41–48 in X, 5–8 between male pores; first

dorsal pore in 10/11; clitellum XIV–XVI, smooth; setae and dorsal pores absent.

Spermathecal pores four pairs in 5/6–8/9, laterally. Distance between paired spermathecal pores about 0.5 body circumference ventrally. Two pairs of papillae present in each of VII and VIII, 1 presetal and 1 postsetal. Some specimens lacking papillae to varying extent; 1 specimen without spermathecal papillae. Some specimens have 1 additional postsetal papilla in VI. Each papilla small, round: distance between paired genital papillae about 0.20 body circumference ventrally. Female pore single, medioventral in XIV. Male pores paired in XVIII, lateroventrally; distance between pores about 0.35 body circumference ventrally. Porophores round or oval on setal line, surrounded by 5-7 circular folds. Genital papillae postsetal, paired in XVII and XIX. One specimen with only the left papilla in XIX. Two specimens with additional papillae in XX: one with a pair, the other with only the right one. Each papilla small, oval, with concave centre.

Preserved specimens reddish brown on dorsum, light yellowish brown on venter.

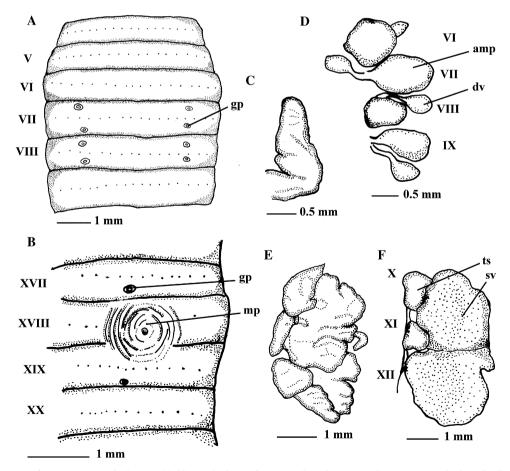


Fig. 3. Amynthas meishanensis, morphology. (A) Ventral view of spermathecal pore region; gp = genital pad. (B) Left male pore area; gp = genital pad, mp = male pore. (C) Right caecum. (D) Right spermathecae; amp = ampulla, dv = diverticulum. (E) Right prostate gland. (F) Left testis sacs and seminal vesicles; ts = testis sac, sv = seminal vesicle.

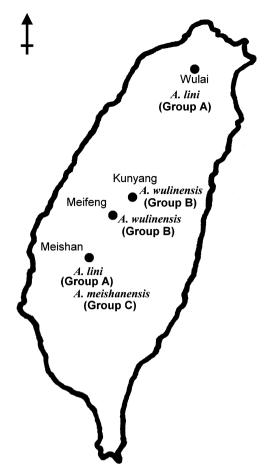


Fig. 4. Collection sites of the Amynthas wulinensis species complex in Taiwan.

Internal characters

Septa 8/9 and 9/10 absent, 10/11–13/14 thickened. Gizzard round in VII–X. Intestine enlarged from XV. Intestinal caeca paired in XXVII, simple, extending to XXIV. Lateral hearts enlarged in X–XIII.

Spermathecae four pairs in VI–IX, with a short stalk about 0.2 mm long and a peach-shaped or oval ampulla about 0.6–1.0 mm long. Spermathecal diverticula present, with a peach-shaped seminal chamber and a straight stalk about as long as seminal chamber. No nephridia on spermathecal ducts. Ovaries paired in XIII, medioventral, close to septum 12/13.

Testes paired in X and XI, small, irregular. Seminal vesicles paired in XI and XII, large. Prostate glands paired in XVIII, large, extending anteriorly to XVII and posteriorly to XX, with a thick duct.

Localities and habitats

Amynthas meishanensis sp. n. inhabits secondary broadleaf forests in central Taiwan, at elevations around 600 m (Fig. 4). It is often found in ditches without water but filled with soil in this mountainous area.

Remarks

Amynthas lini and A. meishanensis correspond to clades A and C, respectively, in the phylogenetic analyses. They are morphologically similar to A. wulinensis s. str.; the

Table 3. Morphological comparison of Amynthas wulinensis, A. lini n. sp., and A. meishanensis n. sp. according to original descriptions

	A. wulinensis	A. lini	A. meishanensis
Body length (mm)	63–174	212–254	38–65
Clitellum width (mm)	5.6–6.1 ^a	7.0-9.0	2.7–3.5
Number of segments	70–132	117–129	51–113
First dorsal pore position	11/12	12/13	10/11
Prostomium	Epilobous	Epilobous	Epilobous
Secondary segmentation	3 annuli after VI	3 annuli after VI	1 annulus for all segments
Setae			
V	ND	20–26	27–31
VII	28–45	33–45	35–42
X	ND	49-57	41–48
XX	33–69	ND	ND
Between male pores	8–13	8–17	5–8
Number of spermathecae	4 pairs	4 pairs	4 pairs
Position of testes	10 and 11	10 and 11	10 and 11
Position of seminal vesicles	11 and 12	11 and 12	11 and 12
Position of prostate glands	15–20	17–21	17–20
Position of preclitellum papillae	Absent	Absent or 7–10	Absent or 6–8
Position of postclitellum papillae	17, 19 and 20	17, 19 and 20	17, 19 and 20

ND: no data.

^aData from reported non-type material, not available in original description.

three species form the A. wulinensis species complex. They are holandric, octothecate earthworms belonging to the corticis species group of the genus Amynthas (Sims and Easton 1972). Most morphological characters do not differ between the three species (Table 3). However, the numbers of setae between the male pores are generally higher in A. lini and A. wulinensis than in A. meishanensis, but they still overlap among the three species. The male pores of the three species are quite similar, but those of A. meishanensis have more circular folds around the porophores, and the associated genital papillae are comparatively smaller (Figs. 2 and 3). Spermathecal papillae are often present in A. lini and A. meishanensis, but they are absent in A. wulinensis. The arrangement of these papillae is more regular in A. meishanensis than in A. lini, and their positions differ (Table 3). Body sizes, including body length and clitellum width, also differ among the three species. A. lini, a species locally called 'snake earthworms' because of its large body size, is larger than the other two species; A. meishanensis is slightly smaller than A. wulinensis. In live specimens, the striped or banded appearance of A. lini is a remarkable feature, making the species easily distinguishable from the other two species.

Discussion

Although included in character descriptions of earthworms, body size and colouration are seldom regarded as species-diagnostic characters because they tend to vary intraspecifically (Gates 1972; Blakemore 2002). The use of these two characters in earthworm taxonomy is also compromised of the procedures of sample processing. Because earthworms are soft-bodied, different concentrations of alcohol used in sample anesthetisation cause different degrees of muscle contraction or relaxation, which obviously affects the measurable body length and width of earthworm specimens. Even when routinely using 10% alcohol, the different sample treatments before fixation or preservation also cause differences in body biometry. On the other hand, the use of 5-10% formalin for fixation and 70% alcohol for preservation both result in the decolouration of earthworm specimens. In addition, with these treatments, the transparent portions of specimens usually become whitish; and the so-called 'alcoholic browning' occurs in specimens after long-term preservation in alcohol (Gates 1972). These make most specimens look similar in colour, whether they are conspecific or not.

The three species of the *A. wulinensis* species complex, *A. lini* n. sp., *A. meishanensis* n. sp., and *A. wulinensis*, are morphologically similar. For this reason, they were formerly treated as a single species. However, after comparing larger numbers of specimens, we find that the combined use of genital papillae, body size and

colouration can lead to correct identifications within the *A. wulinensis* species complex. Actually, when the target specimens have no preclitellum genital papillae, body size and colouration are the most important characters for distinguishing between *A. wulinensis* and *A. lini*.

The use of body size and/or colouration as diagnostic characters is rare in earthworm taxonomy. However, a famous case is the distinction between E. fetida and E. andrei. Concerning morphology, E. fetida has a striped or banded appearance whereas E. andrei is uniformly reddish, but aside from this difference in colouration, the two species are indistinguishable (Domínguez et al. 2005). This similarity caused a 30year discussion on whether or not the two colour morphs are conspecific. Now it is clear that they are indeed two different species (Roch et al. 1980; Valembois et al. 1982; Domínguez et al. 2005; Pérez-Losada et al. 2005), and that many specimens treated as E. fetida in ecotoxicological studies or organic waste management actually represent E. andrei or a mixture of both species (Domínguez et al. 2005). Besides the instances of the A. wulinensis species complex and E. fetida/E. andrei, we believe that body size and colouration may also be useful in species identification in other species complexes, in which similar species are hard to differentiate with the traditionally used characters.

Undoubtedly, standardised methods of comparing body size and colouration for earthworms are currently unavailable. When making body size comparisons, most earthworm taxonomists use body length and/or width, which can vary even within a specimen. Comparisons of colouration are usually based on some imprecise terms of colour description, such as 'bluish brown', 'reddish grey', etc. These imprecise treatments of body size and colouration make it difficult to compare these characters reported from various published studies and to use them for species identification. For both purposes, further studies on developing standardised protocols to measure and record body size and colouration are necessary.

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